Detection of ESBL and carbapenemases in Gramnegatives

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- Background
- ESBL detection methods
- CRE detection methods
- Q&A
- Post-webinar assignment





- ESBL: Extended-Spectrum Beta-Lactamase
- AmpC: Beta-lactamase of the AmpC-type
- Carbapenemase

...ase = suffix to form enzyme name (added to the end of the substrate name)





- Which bacteria carry beta-lactamases?
 - Found in many types of bacteria mainly gram-negative. Some examples are:

| Enterobacterales | | Non-Enterobacterales | Gram-positive | |
|------------------|-----------------------|------------------------|------------------------|--|
| • | Escherichia coli | Pseudomonas aeruginosa | • Staphlococcus aureus | |
| • | Salmonella enterica | Pseudomonas putida | -MRSA | |
| • | Klebsiella pneumoniae | • Acinetobacter spp. | • Others | |
| • | Klebsiella oxytoca | Neisseria gonorrheae | | |
| • | Citrobacter freundii | Haemophilus influenzae | | |
| • | Enterobacter spp. | | | |
| • | Proteus mirabilis | • Campylobacter spp. | | |
| • | Serratia marcescens | | | |
| | | | | |





• Beta-lactam antibiotics:

| Class | Group | Effect | Examples (generation) |
|--------------|--|--------------------|---|
| | Penicillins | Narrow spectrum | Benzylpenicillin (1. gen) Oxacillin (2. gen) |
| | | Extended spectrum | Amoxicillin (3. gen) Piperacillin (4. gen) |
| | Cephalosporins | | Cefalotin (1. gen)/ Cefoxitin (2. gen) |
| Beta-lactams | (Cephamycins) (Cephems) | Extended spectrum | Cefotaxime, Ceftazidime (3. gen) |
| | | | Cefepime (4. gen) |
| | | MRSA | Ceftaroline (5. gen) |
| | | MDR | Cefiderocol (Others - siderophore) |
| | Monobactams | | Aztreonam |
| | Combinations with <u>beta-lactam</u> inhibitors | ESBL | Cefotaxime or ceftazidime with <u>clavulanic acid</u> |
| | Carbapenems | MDR | Meropenem |





- Carbapenem antibiotics:
 - Carbapenems are β -lactam antibiotics similar to penicillin
 - meropenem, imipenem, ertapenem, doripenem
 - Carbapenems are highly resistant to most β-lactamases
 - Broad spectrum of activity
 - Both Gram-positive and Gram-negative bacteria

| | Strep spp. & MSSA | Enterobacteriales | Pseudomonads, Acinetoobacter spp | Anaerobes |
|-----------|-------------------|-------------------|-------------------------------------|-----------|
| Meropenem | + | + | + | + |
| Imipenem | + | + | + | + |
| Ertapenem | + | + | Limited activity | + |
| Doripenem | + | + | + | + |





- Which bacteria carry Carbapenemases?
 - Carbapenemases are mainly found in Gram-negative bacteria

| Enterobacteriales | | Non-Enterobacteriales | | | | |
|-------------------|------------------------|------------------------|--|--|--|--|
| • | Klebsiella pneumoniae | Pseudomonas aeruginosa | | | | |
| • | Klebsiella oxytoca | Pseudomonas putida | | | | |
| • | Escherichia coli | • Acinetobacter spp. | | | | |
| • | Citrobacter freundi | | | | | |
| • | Enterobacter aerogenes | | | | | |
| • | Enterobacter cloacae | | | | | |
| • | Enterobacter gergoviae | | | | | |
| • | Proteus mirabilis | | | | | |
| • | Salmonella enterica | | | | | |
| • | Serratia marcescens | | | | | |





- Carbapenemase-producing *Enterobacteriaceae* (CPE):
 - Gram negative bacteria that can live in human and animal gut
 - Klebsiella spp., E. coli, Enterobacter spp.
 - Involved in a variety of infections
 - Bacteremia, urinary tract infections, intra-abdominal infections and pneumonia
 - High mortality rates associated with CPE infections as high as 57%*
 - Urgent need of new treatment strategies for treating CPE infections





- Screening methods
 - Broth/agar dilution or disk diffusion method
 - Selective media
- Phenotypic confirmatory methods
- Genotypic confirmatory methods



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ESBL screening methods

| Method | Antibiotic | Conduct ESBL-testing if | | |
|-------------------------------------|--|--|--|--|
| Broth or agar dilution ¹ | Cefotaxime/ceftriaxone AND Ceftazidime | MIC >1 mg/L for either agen | | |
| | Cefpodoxime | MIC >1 mg/L | | |
| Disk diffusion ¹ | Cefotaxime (5 µg) or Ceftriaxone (30 µg) AND Ceftazidime (10 µg) | Inhibition zone <21 mm Inhibition zone <23 mm Inhibition zone <22 mm | | |
| | Cefpdoxime (10 µg) | Inhibition zone <21 mm | | |

¹ With all methods test either (i) cefotaxime or ceftriaxone AND ceftazidime OR (ii) cefpodoxime alone.

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Figure 1. Algorithm for phenotypic detection of ESBLs



¹ If cefoxitin has been tested and has an MIC >8 mg/L, perform cefepime+/- clavulanic acid confirmation test
² Cannot be determined as either positive or negative (e.g. if a gradient diffusion strip cannot be read due to growth beyond the MIC range of the strip or there is no clear synergy in combination-disk and double-disk synergy tests). In confirmation with cefepime +/- clavulanic acid is still indeterminate, genotypic testing is required.

EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance. Version 2,0, July 2017



- Culture media in-house (with cefotaxime and/or ceftazidime)
- Commercial ready-to-use media plates
- Chromogenic media (with cefpodoxime)





ESBL Phenotypic confirmatory methods

- 1. Combination disk test (CDT)
- 2. Double-disk synergy test (DDST)
- 3. Gradient method Etest ESBL
- 4. Broth microdilution



Combination disk test (CDT)

- Disk containing cephalosporin alone (cefotaxime, ceftazidime, cefepime) and in combination with clavulanic acid
- Compare inhibition zones

ceftazidime 30µg

cefotaxime 30µg

ceftazidime 30µg + clavulanic acid 10µg

cefotaxime 30µg + clavulanic acid 10µg

positive if ≥5 mm increase in inhibition zone with clavulanic acid



Double-disk synergy test (DDST)

- Disk containing cephalosporin (cefotaxime, ceftazidime, cefepime) is applied next to disk with clavulanic acid (amoxicillin-clavulanic acid)
- Shorter distance between disks 20mm center-to-center



*: May be reduced to 15mm or expanded to 30mm if high or low resistance level. EUCAST uses lower disk content → re-evaluation needed!

 positive if inhibition zone around cephalosporin disk is enhanced in the direction of the disk with clavulanic acid



Gradient method – Etest ESBL

- 2-sided strips containing gradients of cefotaxime, ceftazidime or cefepime either alone or in combination with clavulanic acid
- according manufacturer's instruction



MIC ratio: CT/CTL = 1.5/0.047 = 32

• positive if MIC ratio \geq 8 (MIC reduced by three doubling dilution steps)



Gradient method – Etest ESBL

- according manufacturer's instruction
- 2-sided strips containing gradients of cefotaxime, ceftazidime or cefepime either alone or in combination with clavulanic acid



- positive if MIC ratio \geq 8 (MIC reduced by three doubling dilution steps)
- or phantom zone/deformed ellipse is present
- indeterminate if growth beyond the MIC range of the strip



False negative results

• ESBL presence masked due to high-level expression of AmpC β-lactamases

 clear resistance to 3rd generation cephalosporins + to cephamycins e.g. cefoxitin MIC > 8mg/L (except ACC AmpC β-lactamases)

Perform **confirmatory test** with **cefepime** as indicator cephalosporin or use **cloxacillin**

Cefepime: usually not hydrolyzed by AmpC β-lactamases → CDT, DDST, Etest, broth microdilution Cloxacillin: good inhibitor of AmpC enzymes → CDT with disk containing cloxacillin or CDT/DDST with agar supplemented with cloxacillin

 ESBL presence masked by carbapenemases such as MBLs or KPCs (not OXA's) and/or severe permeability defects



False positive results

- Klebsiella oxytoca with hyperproduction of chromosomal K1 or OXA-like βlactamases
- Proteus vulgaris, Citrobacter koseri, Kluyvera spp due to chromosomal βlactamases inhibited by clavulanic acid
- Strains with hyperproduction of SHV-1, TEM-1 or OXA-1-like broad-spectrum β-lactamases combined with alterered permeability



ESBL Genotypic confirmatory methods

- PCR (and sequencing) → epidemiological information is needed to decide which genes to target
- Microarrays → limitations in the number of resistance genes targeted
- WGS \rightarrow well curated and complete databases are needed



ESBL Genotypic confirmatory methods

- Beta-lactamases are numerous!
- ESBLs
 - CTX-M
 - TEMSHV
 - (OXA)
 -
- AmpCs
 - cAmpC

 - ACCDHA
 - FOX
 -

| Correct nomenclature: | | | | | |
|------------------------|---------|--|--|--|--|
| Gene | Protein | | | | |
| bla _{CTX-M-1} | CTX-M-1 | | | | |
| bla _{CMY-2} | CMY-2 | | | | |
| etc | etc | | | | |

- Each type of enzyme is further divided into variants: E.g. CTX-M-1, CTX-M-2...
- You can find a database of beta-lactam genes here: <u>http://bldb.eu/</u>



CRE Detection methods

- Screening methods
 - Broth/agar dilution or disk diffusion method
 - Selective media
- Phenotypic confirmatory methods
- Genotypic confirmatory methods



Broth/agar dilution or disk diffusion method

| | N | 1IC breal | <point (mg="" l)<="" th=""><th colspan="3">Zone diameter breakpoint (mm with 10µg disks</th></point> | Zone diameter breakpoint (mm with 10µg disks | | |
|-----------|-----|-----------|--|---|----|--|
| | S≤ | R> | | S≥ | R< | |
| Ertapenem | 0.5 | 0.5 | | 23 | 23 | |
| Imipenem | 2 | 4 | | 22 | 19 | |
| Meropenem | 2 | 8 | | 22 | 16 | |



Broth/agar dilution or disk diffusion method

| | MIC breakpoint | | | nt (mg/L) Zone diameter br with 10μ | | |
|-----------|----------------|-----|----------------|--|----|---------|
| | S≤ | R> | cut-off | S≥ | R< | cut-off |
| Ertapenem | 0.5 | 0.5 | → 0.125 | | 25 | ∢-25 |
| Meropenem | 2 | 8 | > 0.125 | 22 | 16 | < 28* |

Use **ECOFF** values

Ertapenem: excellent sensitivity but poor specificity, especially in species such as *Enterobacter spp.* (relative instability to ESBLs and AmpC in combination with porin loss

 \rightarrow not recommended for routine use

Imipenem: the separation between the wild-type and carbapenemase-producers is relatively poor

 \rightarrow not recommended for use as a stand-alone screening test compound

Meropenem: best balance of sensitivity and specificity

* Isolates with 25-27 mm only need to be investigated for carbapenemase production if they are resistant to piperacillin/tazobactam and/or temocillin

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CRE screening





CRE screening methods

- Culture media in-house (with meropenem)
- Commercial ready-to-use media plates
- Chromogenic media





• meropenem (10µg) +/- various inhibitors

Table 2. Interpretation of phenotypic tests (carbapenemases in **bold type**) by diffusion methods with disks or tablets. The exact definitions of synergy are provided in package inserts for the various commercial products.

| B-lactamase | Synergy obse (mm) with 10 | Temocillin MIC >128 | | | |
|------------------------|------------------------------|------------------------|----------|-----|------------------------------------|
| | DPA/EDTA | APBA/PBA | DPA+APBA | CLX | mg/L or zone diameter <11 mm |
| MBL | + | - | - | - | Variable ¹ |
| КРС | - | + | - | - | Variable ¹ |
| MBL + KPC ² | Variable | Variable | + | - | Variable1 |
| OXA-48-like | - | - | - | - | Yes |
| AmpC + porin loss | - | + | - | + | Variable ¹ |
| ESBL + porin loss | - | - | - | - | No |

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• meropenem (10µg) +/- various inhibitors



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Meropenem 10µg

Meropenem 10µg + DPA 1000µg Meropenem 10µg + cloxacillin 750µg

Meropenem 10µg + APBA/PBA 600µg



Interpretation criteria?

Compare inhibition zone diameter of meropenem with meropenem + inhibitor



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- rapid (2 hours), cheap, high sensitivity and specificity biochemical test
 - change of phenol red color (red to yellow) after hydrolysis of imipenem by carbapenemase





Carbapenem Inactivation Method (CIM)

Low-cost alternative to the CarbaNP test





CRE Genotypic confirmatory methods

- Multiplex PCR
- Real-time PCR
- Cepheid GeneXpert Carba-R
 - Detection and differentiation of KPC, NDM, VIM, IMP-1, and OXA-48 in 48 minutes
- Whole-genome sequencing



CRE Genotypic confirmatory methods

- Sequencing of entire gene or genome might be needed to differentiate between variants and derivatives
 - For example, to differentiate between OXA-48 (true carbapenemase) and OXA-163 (weak carbapenemase)
- Screening of sequenced genome against a database of known carbapenemase genes using bioinformatics tools
 - Continuously updated tools and databases such as ResFinder, AMRFinderPlus, and CARD
- Both known and unknown variants can be detected with WGS







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Post-webinar assignment

- All participants in this webinar will receive an email with a link and brief instructions for the assignment
- 6 cases. Case background provided, and you'll answer related questions.
- Deadline for submitting results: 30th of May 2024
- Should you have any questions or need further clarification, feel free to reach out to us at eqasia@food.dtu.dk







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